Dog Manual Ejaculation

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Book Descriptions:

Dog Manual Ejaculation

This is a hands on demonstration for dog breeders to learn howto stimulate the dogs penis, slip a collection hood over the penis and then gather the sperm rich ejaculate. Please enable JavaScript to watch this video. Jumpstart your career with our Premium AtoZ Microsoft Excel Training Bundle from the new Gadget Hacks Shop and get lifetime access to more than 40 hours of Basic to Advanced instruction on functions, formula, tools, and more.Can you say beastiality! Gross! I never would have thought to use a collection hood, I was using less conventional methods0. I thank ppl for showing stuff like this to help others out and now we will have a litter thanks to this movie. Whats cruel is not giving your dog a hand from time to time lol. Imagine spending the rest of your life horny and having no hands to stimulate yourself and no lady dog to take care of you. That would suck lol! Its nothing special, but you must have the skills. I know Boxer is a high energy dog, but he does nothing but chase my female trying even when not in heat. Who is the pervert now Very Professional. Never heard of cryopreservation of semen. But insemination on human is REALLY gross guess how they perform it. This is a very essential bit of information a dog breeder needs to know. It may seem discusting to narrow minded immature individuals with no education in the dog breeding realm but i do assure you this is an educational video for dog breeders. If you are not breeding dogs why are you here viewing this. Sick curiosity of a twisted mind can make anything dirty. I will leave it to the veterinarians who do this routinely We do not sell personal information to 3rd parties. The usual recommendation is neither to use a dog for breeding nor to collect semen more often than every other day. Many of the spermatozoa in an ejaculate come from storage in the epididymis. Whenever a dog ejaculates, many of those stored spermatozoa are released.http://acecaalcoy.com/userfiles/casio-dl-250la-user-manual.xml

- **dog manual ejaculation.**

If the dog is used more frequently than every other day, the number of spermatozoa in the ejaculate declines with each collection as that reserve is depleted. Most dogs also show a decrease in libido if they are used or collected very frequently. 2 What is a “rusty load”. A rusty load is an ejaculate containing many old spermatozoa from storage, many of which are misshapen and have poor motility. If semen is collected from a dog that has not been used for a while and many of the spermatozoa appear abnormally shaped, semen should be collected again before that dog is evaluated critically. 3 How do you get a semen sample from a dog. Semen is collected from dogs by manual ejaculation. The dog does not need to be sedated. You will get a betterquality sample if there is a teaser bitch present, but often you can get an adequate sample with no teaser present. The dog’s penis is stimulated, and as he becomes erect, the penis is directed toward a collection vessel. I like to use a rubber collecting cone that fits over the penis and prevents loss of the semen sample onto the floor, but you can collect semen into a cup. I. SEMEN COLLECTION Manual ejaculation of dogs is a vital component of a breedingsoundness examination, the definitive diagnostic test for male dogs presenting for subinfertility or infertility, and the technique used to collect semen for artificial insemination or semen preservation. Electroejaculation, in which a rectal probe is used to stimulate ejaculation neurologically, is not performed in domesticated dogs. Semen can be collected into any container. I have heard of people using cups, sandwich bags, and other household items. Any container used must be very clean and contain no soap residues; any contaminant may kill spermatozoa collected into that vessel.http://detaycopymatbaa.com/userfiles/casio-dr-t120-manual.xml

I prefer to use a rubber collecting cone with a clear plastic centrifuge tube attached to the end

Figure 231 because it is a selfcontained unit that does not allow semen to be spilled or ejaculated outside of the collecting instrument and because it best mimics natural service, presumably yielding a better quality ejaculate. Again, the equipment must be kept scrupulously clean. Rubber should not be cleaned with soap at any time because soap residues easily adhere to the rubber and may kill spermatozoa subsequently collected with that equipment. Figure 231 Rubbercollecting cone for canine semen collection. Most male dogs can be collected in the absence of a teaser bitch. However, it has been demonstrated that the libido of the male and quality of the semen are improved in the presence of a teaser bitch, especially if she is in heat. Some very experienced male dogs require not just a bitch in heat but one that is at the optimal time for breeding. If a teaser bitch is not available, there are several techniques that can be used to entice the male 1 use of a commercial product mimicking the pheromones produced by a bitch in heat, 2 allowing the male to sniff at swabs collected from an estrous bitch and frozen until needed, and 3 administration of a drug that stimulates contraction of the smooth muscle in the epididymis. In my experience, the first two techniques have not proven very successful. The last technique involves administration of prostaglandin Lutalyse 10 minutes before attempted semen collection. This technique has been demonstrated to increase number of spermatozoa in the ejaculate to the same extent as does the presence of an estrous teaser bitch. The floor should be a nonslip type. If semen collection is to be performed with any regularity, a consistent surface, such as a rug, should be used. This provides secure footing and trains the dog as to what is expected of him when the rug is present.

If a teaser bitch is used, a handler should hold her with her hindquarters on the rug. If the bitch is not in heat, she should be muzzled or otherwise securely restrained. The male is allowed to investigate her hindquarters and to mount if he so wishes. The bulbus glandis is palpated through the prepuce. The area over the bulbus glandis is manipulated briskly and enthusiastically. As soon as the penis starts to become erect, the prepuce is pushed behind the engorging bulbus glandis. This is best accomplished by using the rubber collecting cone to push the prepuce back while “squirting” the penis out of the prepuce with the other hand. A betterquality ejaculate usually is obtained if the prepuce is pushed proximal to the erect bulbus glandis. Once the rubber collecting cone is advanced completely proximal to the bulbus glandis, the penis between the bulbus glandis and body wall is encircled tightly with the fingers. This mimics the tie. At this point, most dogs thrust vigorously and ejaculate a clear presperm fraction and then the cloudy spermrich fraction of the ejaculate Figure 232. The dog stops thrusting and steps down. He may try to step over the collector’s arm. If the bulbus glandis is completely free from the prepuce, the collector should swing the dog’s penis 180 degrees in a horizontal plane and continue to hold. The final fraction, the prostatic fraction, is ejaculated as distinct pulses of clear fluid that will be visible filling the tube attached to the collecting cone and will be palpable in the hand holding the collecting cone and penis. The anus contracts rhythmically as prostatic fluid is ejaculated. The amount of prostatic fluid collected depends on the purpose of the semen collection Table 231 . At the conclusion of semen collection, the penis is released and the rubber collecting cone gently peeled off the penis.

Allow the male to lick at the penis, walk him away from the collection area, or gently massage the engorged penis with towels moistened with cold water not ice to hasten detumescence of the penis. Ensure that the penis is completely flaccid and within the prepuce and that the prepuce is not rolled in on itself before kenneling the male dog. Figure 232 Semen collection in the dog. Table 231 Prostatic Fluid Collection During Manual Ejaculation Purpose of Semen Collection Amount of Prostatic Fluid to Collect Breeding soundness exam complete semen evaluation One to two good “squirts,” enough to ensure the entire second “spermrich” fraction has been collected Semen collection for culture and cytology testing or other diagnostic tests prostate disease, subfertility, or infertility As above or enough to provide sufficient volume for tests to be conducted Artificial

insemination with fresh semen see Chapter 21 Dependent on size of bitch to be inseminated. For bitches weighing less than 10 lb, total semen volume should be 1.5 to 3 mL; for bitches weighing 10 to 50 lb, 3 to 5 mL; and for bitches weighing more than 50 lb, 5 to 8 mL Semen to be chilled and shipped overnight One good “squirt”—amount of prostatic fluid collected should be minimized as incubation of spermatozoa with the dog’s prostatic fluid is associated with decreasing motility of those spermatozoa Semen to be frozen One good “squirt,” enough to ensure the entire spermrich fraction has been collected. Sample will be centrifuged and the spermatozoa removed for further processing II. SEMEN EVALUATION The components of a complete semen evaluation include measurement of volume, assessment of color, subjective assessment of percentage progressively motile spermatozoa, determination of concentration and total number of spermatozoa in the ejaculate, and assessment of spermatozoal morphology shape. A. VOLUME Volume is not an indicator of semen quality because it is dependent on the amount of prostatic fluid collected.

However, the value for volume must be recorded to allow determination of the total number of spermatozoa in the ejaculate. A normal value for volume has been reported at 1 to 30 mL Figure 233. Figure 233 Volume assessment. B. COLOR Color is a superficial indicator of semen quality and possible contamination of semen. Semen should be milky or opalescent. All milky fluid collected should be evaluated microscopically because occasionally dogs will ejaculate fluid that contains no spermatozoa but has many fat droplets that give it a milky appearance. Yellow is indicative of urine contamination. Brown or red indicates that blood is present in the semen. Brown discoloration is due to old blood and usually is associated with prostate disease. Red discoloration is due to frank blood and may be associated with prostate disease or with trauma to the penis. Occasionally young male dogs bleed from the surface of the penis the first time they become erect because small blood vessels rupture on the engorging penis. Green discoloration may be seen in animals with reproductive tract infection. Finally, clear fluid indicates lack of spermatozoa in the ejaculate azoospermia Figure 234. Figure 234 Color assessment. C. MOTILITY The percentage of progressively motile spermatozoa is assessed by evaluation of a drop of semen on a glass slide, examined microscopically at 40 to 100.This is a very subjective assessment of what percentage of spermatozoa are moving forward. Some investigators evaluate total motility, progressive motility those moving forward, and speed of motile spermatozoa. Normal percentage progressively motile spermatozoa is 70% or greater. Log In or Register to continue. Semen collected for insemination can be used fresh, or can be cooled and shipped to another location. Canine semen can also be frozen, allowing long term storage. Another indication for collecting semen is to obtain prostatic fluid for culture or cytology in cases of suspected prostatic disease.

However, use of a bitch will almost certainly expedite the procedure and allow more sperm to be harvested. Ideally the teaser should be in estrus, but considering the length of the canine cycle, that is often difficult to arrange, and a friendly, nonestrus bitch will often serve the purpose. If a bitch is used, she should be controlled with her rear quarters facing the male.Most failures arise because the male is shy or otherwise intimidated. It helps to perform the collection on a nonslip surface such as a carpet. If the male appears nervous or this is his first time, a teaser bitch may help considerably.A latex collection cone with an attached tube is commonly used. As an alternative, some people prefer to use a disposable baby bottle liner. Both collection tools are shown.The basic process is conducted in the following series of stepsMost dogs stop thrusting as they begin to ejaculate. Semen can be collected in this manner, but not as easily. If this happens, simply take the male a distance away to let him calm down, then try again.By using a clear plastic collection tube, the delivery of these three fractions is easily monitored.In most cases, collection is stopped as soon as this fraction is recognized. Males may successfully breed bitches prior to sexual maturity but they will not achieve maximal fertility or daily sperm output until mature. The normal male can breed once every 2 5 days and maintain daily sperm output. A complete breeding

soundness examination in the dog includes historytaking, general physical examination, reproductive system examination, libido determination, semen collection and evaluation, hormonal evaluation, and prostatic examination and testing for disease eg. Brucellosis.

Semen collection In addition to artificial insemination breeding, collection of semen is indicated for evaluation in conjunction with a breeding soundness examination, in the diagnostic workup of potentially subfertile or infertile dogs, in the diagnostic workup of reproductive tract disease infectious, degenerative or neoplastic disease, or for shortterm freshchilled extended semen or long term frozen semen storage of gametes to be used in the future. When very young 12 years sires are used for breeding, the American Kennel Club AKC requires documentation, including a semen evaluation, of the males ability to sire a litter. In the typical setting, ejaculation in the dog is a voluntary process and cannot be forced or rushed. Some males may not require any external stimulus beyond manual massage to attain erection. If a teaser bitch is not available, many dogs will respond favorable to swab scent from an estrous bitch. He may sniff or lick her external genetalia or rear limbs. He may or may not mount the bitch. Some males will achieve erection without any manual manipulation. Other males require brisk massage of the bulbus glandis through the prepuce to elicit erection. As soon as erection begins, the prepuce is moved proximally such that the entire prepuce is proximal to the exposed bulbus glandis. Failure to reposition the prepuce at the correct time may result in an inability to slide the prepuce proximally over the bulbis glandis. The collection process may then become painful to the dog as the skin of the prepuce tightens over the engorged bulbis glandis. It is important to roll over the edge of the cone such that the edge of the cone does not come into direct contact with penis. During this period, the first and second seminal fractions are ejaculated. The first or presperm fraction is of prostatic origin and is clear. The third fraction is comprised of prostatic fluid and is normally clear.

During third fraction collection, the dog may try to step over the collectors arm. If all three fractions of the ejaculate need to be collected, it is advisable to collect the fractions into separate receptacles. The dog often will continue to ejaculate pulses of prostatic fluid. Detumescence of the penis may require several minutes and often takes a similar amount of time as would recovery from a natural mating. Occasionally the prepuce rolls inward as penile engorgement wains leaving the unprotected glans of the penis vulnerable to dessication and trauma. As much as a fourfold increase in spermatozoa number can be realized when using a teaser bitch. Semen handling Sperm are motile and vigorous cells, but are also fragile and susceptible to damage and demise by environmental conditions. When collecting and handling semen it is critical to avoid exposing sperm cells to two types of insults toxic chemicals and thermal insult. Keeping collection equipment clean and disinfected is important, but soap and disinfecting chemicals are quite potent spermicides. For this reason and for their convenience of use, disposable polypropylene collection cones are often preferred over their latex counterparts. When using latex cones, take great care to rinse thoroughly deionized water to remove all soap and disinfectant residue. It is usually best to use new, sterile collection tubes. Finally, be certain to lubricate collection cones with a sterile lubricant known to be nontoxic to sperm. If the tube of lubricant is not clearly labeled as being nonspermacidal then you cannot presume that it is safe to use. Sperm are sensitive to both heat and cold. Short periods of exposure to temperatures just a few degrees above body temperature will usually kill large numbers of sperm. To avoid thermal stress, the collection cone should be prewarmed to body temperature.

Additionally, microscope slides, coverslips, stains, extenders and pipets used to handle and examine sperm are best maintained on a warming plate prior to use. If you are performing a large number of analyses, a heated microscope stage is also a valuable piece of equipment. A large number of extenders have been developed and are commercially available for use in short term and long term storage of semen. They are similar in having an energy source eg.Tris or citrate and a source of protein eg.Semen evaluation Putting the evaluation into perspective Ultimately, the goal of any

semen evaluation is to predict the likelihood for a dog to be successful in siring a litter. Routine semen evaluation in the dog includes determination of semen volume, color and pH, spermatozoa motility, velocity, concentration, total number of spermatozoa in the ejaculate, and sperm cell morphology. In humans, The World Health Organization WHO has developed strict criteria for semen collection and evaluation. No such criteria have been adapted to any of the domestic species, including the canine, beyond the guideline that a dog should not be condemned on the results of one semen evaluation. Male factor infertility accounts for up to 50% of failed pregnancy attempts in humans. Compared to human medicine, little is known in canine medicine regarding specific findings on semen evaluation and their correlation with fertility. No estimates have been reported for the canine, but experiential evidence supports the notion that male factor infertility does account for a significant portion of conception failure, early embryonic death and small litter size. Veterinarians routinely perform semen analysis on dogs. Unfortunately, there is little data associating the parameters measured with the issues practitioners really need to evaluate, i.e. testicular function, fertilizing capability of spermatozoa, and likelihood that puppies will develop normally.

With the current methodology, semen analysis may only be reliably predictive of fertility if the semen quality is either very good or very bad. A big problem is that in vitro analysis.Results of tests that may be performed in a semen evaluation are influenced by sample collection technique and timing, concentration of spermatozoa in the sample, amount of time from sample collection to evaluation, temperature at which the sample was held, equipment used, and many other factors, not the least of which is the subjectivity of the evaluator. Another impediment to standardization of semen evaluation in the canine relates to the enormous variability present as a result of breed. Along with the obvious extremes in testicular size, there may be other significant influence of breed on semen parameters. Acknowledging the problems encountered and limitations inherent with making functional conclusions on the basis of in vitro testing, multiple parameters are used in performing a semen evaluation. Color and appearance Color evaluation is subjective but informative. The turbidity or opacity of semen provides a rough indication of concentration. Commonly, the turbidity of a sample is graded on a subjective scale of 0 to 5, with 5 being the most opaque as in the whole milk. A clear sample contains no spermatozoa. Although cloudy or milky samples almost always contain spermatozoa, they must still be checked microscopically because occasionally a dog with azoospermia will shed excessive numbers of fat droplets into the sample, giving the appearance of normal semen. Yellow semen is indicative of urine contamination and is also seen in humans with icterus or after ingestion of certain vitamins. Brown discoloration is indicative of old digested blood and red discoloration is indicative of fresh blood. The most common causes of hemospermia in dogs are prostatic disease and penile trauma.

In humans, hemospermia also may be associated with urinary calculi, sexually transmitted diseases such as syphilis and gonorrhea, spermatocele and hydrocele, and treatment with anticoagulant medications.Values for normal pH in nonfractionated canine semen vary from 6.4 to 6.8. Seminal pH may change in the presence of disease, such as prostatitis, or if the semen sample is contaminated with urine. There are no published data comparing pH analysis techniques for canine semen or specifically describing the clinical value of pH measurement. Alterations in pH may affect sperm longevity and motility. Decreases in prostatic fluid pH are common with prostatic disease. Alterations in pH may occur with use of excessive amounts of lubricant or improper cleaning and disinfection of collection equipment. Volume Dog semen is ejaculated in three fractions. The first, presperm fraction is small in volume less than 5 mls and contains few to no spermatozoa. The presperm fraction is believed to cleanse the urethra of contaminants urine, bacteria and cellular debris prior to ejaculation. The presperm is not usually collected. The second, spermrich fraction comes from the epididymes and testes. The volumes of the first and third fractions are variable. In particular, the volume of the third fraction is controlled by the person collecting the sample, as they

can choose to collect more or less of the cellfree prostatic fluid. Volume is not an indicator of semen quality in dogs. However, the measurement of semen volume is important in the calculation of total number of spermatozoa in the sample. Motility Sperm motility is essential for fertilization because it allows or at least facilitates passage of the sperm through the zona pellucida. Without technologic intervention, a nonmotile or abnormallymotile sperm is not going to fertilize. Hence, assessing the fraction of a sperm population that is motile is perhaps the most widelyused measure of semen quality.

For canine semen, motility is better maintained if samples are kept at room temperature than at body temperature. Warmer temperatures increase sperm cell metabolism. Delay in evaluation of warmed samples may result in errors due to sperm cell energy depletion. Rapid temperature fluctuations should be avoided. Ambient temperature may affect the motility assessment of a sample if the evaluation room is excessively hot or cold. Percentage of progressively motile spermatozoa from a given dog is not affected by frequency of semen collection. In evaluating motility, sperm cells are classified as immotile, progressively motile or nonprogressively motile. Both total and progressive motility are determined and expressed as a percentage of 100. Sperm cells showing motility of any kind are included in the calculation of total motility. A progressively motile spermatozoan moves forward in an essentially straight line, whereas a nonprogressively motile sperm cell moves, but with an abnormal path, such as in tight circles tail chasing, fish flopping, and when normal movement is prevented by sperm head agglutination. In the dog, the normal percentage of progressively motile spermatozoa is 70% or greater. Progressively motility is positively correlated with normal morphology in dogs. If the sperm cell is put together normally, then it should move normally. Exposure to rapid or extreme temperature fluctuations, any kind of residue on collection equipment, or the wrong pH or osmolality of an extender can adversely affect motility. Motility is also affected by periods of sexual inactivity males that have not ejaculated for prolonged periods often have poor motility on the first ejaculate, but much better motility for a second ejaculate collected soon thereafter. Manual motility estimates are easy to perform and require minimal equipment.

This commonlyused technique involves placing a sample of semen on a microscope slide, examining it with a microscope and estimating the fraction of the population that is motile. Manual motility estimates are subjective. However, with an experienced evaluator, manual estimates generally provide good estimates of motility. The chief limitation of this technique is its subjective nature. Computer assisted sperm analysis CASA CASA is a technique employing a computerized system that tracks multiple motility parameters. The computer takes video images of the sperm and stores them for analysis. The system recognizes motile from nonmotile sperm and other organic debris by comparing luminosity grayscale intensity and size of the object. There are also preset userdefined thresholds for size and luminosity that help prevent mistaking other cells and debris for nonmotile sperm. While some studies have shown that CASA systems are more accurate than subjective assessment of sperm motility, other studies have found that subjective analysis progressively motility is well correlated with computerbased analysis. Most CASA systems are found in teaching hospitals and research laboratories. Normal values have not been established for the dog. Another potential value of these sperm motility factors is recognition of subpopulations of spermatozoa within the sample that may react differently to cryopreservation or other manipulations of spermatozoa. Concentration and total number Concentration is important because it is used to calculate total sperm number in the ejaculate. Concentration is inversely related to volume collected. Concentration multiplied by volume is the total number of spermatozoa in the ejaculate; total number of spermatozoa is dependent on testicular size. In dogs, normal total number of spermatozoa is greater than 300 million. A general guideline for total sperm number in an ejaculate is 10 million sperm cells per pound of body weight.

Total number of spermatozoa decreases with frequent semen collection, presumably as epididymal reserves are depleted. Normal dogs may ejaculate oligozoospermic or azoospermic samples due to apprehension or pain. In human medicine, apparently azoospermic samples are centrifuged at greater than 3000 g for 15 min and all the fluid systemically examined before that sample is declared completely deficient of spermatozoa. Concentration can be measured by several methods CASA systems, hemacytometer, optical density measurement, and spermatocrit. The most common means of determining sperm concentration is to simply count sperm under a microscope with the aid of a hemacytometer. A hemacytometer is a glass slide onto which a precision grid has been etched. They are sold for counting blood cells, but work equally well for counting sperm. Since the dimensions of the grid squares and depth of sample chamber are known, it is a simple matter to calculate cell concentration. The hemacytometer technique has been reported to be accurate and is considered the gold standard. The coverslip specific to that hemacytometer should be used. Do not use disposable coverslips. Concentration measurement with a hemacytometer is time consuming compared when compared to other methods. A Total number of spermatozoa in the ejaculate can be calculated by multiplying the volume in mls by the concentration number of sperm cells per ml. A guideline for total sperm number in the ejaculate is 10 million sperm cells per pound of body weight. For example, a 60 pound dog should have 600 million sperm cells in his ejaculate. Morphology To evaluate morphology, 100 individual sperm cells are examined for normal shape and structure. The results of a sperm morphology exam are reported as percent normal. Minimally, the size and shape of the head, midpiece and tail are examined. The head contains the DNA and has a cap the acrosome which contains the enzymes that allow zona penetration and fertilization to occur.